

# Primer and Probe Design

# Primer

Good primer design is essential for a successful PCR reaction. There are many factors to take into account when designing the optimal primers for your gene of interest. Here are some tips to consider when designing primers.

1. In general, a length of 18–25 nucleotides for primers is good.
2. Try to make the melting temperature ( $T_m$ ) of the primers between 52°C and 65°C.
3. If the  $T_m$  of your primer is very low, try to find a sequence with more GC content, or extend the length of the primer a little.
4. Aim for the GC content to be between 40 and 60%, with the 3' of a primer ending in C or G to promote binding.

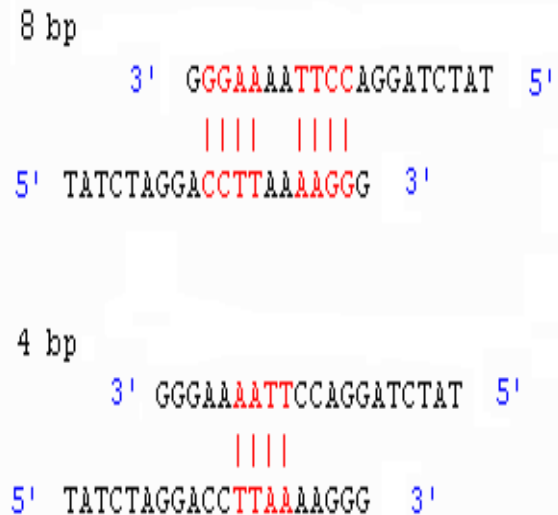
5. Try to avoid regions of secondary structure, and have a balanced distribution of GC-rich and AT-rich domains.
6. Try to avoid runs of 4 or more of one base, or dinucleotide repeats (for example, ACCCC or ATATATAT).
7. Avoid intra-primer homology (more than 3 bases that complement within the primer) or inter-primer homology (forward and reverse primers having complementary sequences). These circumstances can lead to self-dimers or primer-dimers instead of annealing to the desired DNA sequences.

If primers can anneal to themselves, or anneal to each other rather than anneal to the template, the PCR efficiency will be decreased dramatically. They shall be avoided.

### Hairpin



### Self-Dimer



### Dimer



- The common  $T_m$  formulas for calculating the theoretical  $T_m$  of an oligo
- $T_m = 4^{\circ}\text{C} \times (\text{number of G's and C's in the primer}) + 2^{\circ}\text{C} \times (\text{number of A's and T's in the primer})$
- $T_m = 4^{\circ}\text{C}(\text{GC}) + 2^{\circ}\text{C}(\text{AT})$
- $T_a = T_m - (2-5)$

Gene ▾  ✕

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See [FOXP2 forkhead box P2](#)  
 foxp2 in [Homo sapiens](#) [Mus musculus](#) [Rattus norvegicus](#) [All 193 Gene records](#)

### Search results

Items: 1 to 20 of 241

<< First < Prev Page 1 of 13 Next > Last >>

See also 3 discontinued or replaced items.

Name/Gene ID	Description	Location	Aliases	MIM
<input type="checkbox"/> <a href="#">ACOT13</a> ID: 55856	acyl-CoA thioesterase 13 [ <i>Homo sapiens</i> (human)]	Chromosome 6, NC_000006.12 (24667035..24705069)	HT012, PNAS-27, THEM2	615652
<input type="checkbox"/> <a href="#">AES</a> ID: 166	amino-terminal enhancer of split [ <i>Homo sapiens</i> (human)]	Chromosome 19, NC_000019.10 (3052910..3063107, complement)	AES-1-2, ESP1, GRG, GRG5, Grg-5, TLE5, AES	600188
<input type="checkbox"/> <a href="#">Cadm1</a> ID: 54725	cell adhesion molecule 1 [ <i>Mus musculus</i> (house mouse)]	Chromosome 9, NC_000075.6 (47530170..47862280)	2900073G06Rik, 3100001108Rik, AI987920, BI2, Igsf4, Igsf4a, Nec12, RA175, RA175A, RA175B, RA175C, RA175N, ST17, SglGSF, SynCam, Tslc1	
<input type="checkbox"/> <a href="#">CCNC</a>	cyclin C [ <i>Homo sapiens</i> (human)]	Chromosome 6,	CycC	123838

Filters: [Manage Filters](#)

### Results by taxon

Top Organisms [\[Tree\]](#)

- [Homo sapiens](#) (34)
- [Mus musculus](#) (7)
- [Rattus norvegicus](#) (4)
- [Xenopus laevis](#) (3)
- [Taeniopygia guttata](#) (2)
- [All other taxa](#) (191)

More...

### Find related data

Database:

Find items

### Search details

FOXP2[All Fields] AND alive[prop]



polyglutamine tract and is an evolutionarily conserved transcription factor, which may bind directly to approximately 300 to 400 gene promoters in the human genome to regulate the expression of a variety of genes. This gene is required for proper development of speech and language regions of the brain during embryogenesis, and may be involved in a variety of biological pathways and cascades that may ultimately influence language development. Mutations in this gene cause speech-language disorder 1 (SPCH1), also known as autosomal dominant speech and language disorder with orofacial dyspraxia. Multiple alternative transcripts encoding different isoforms have been identified in this gene.[provided by RefSeq, Feb 2010]

Orthologs [mouse](#) [all](#)

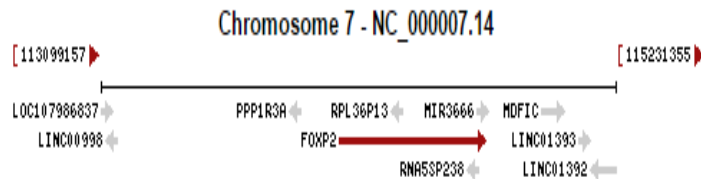
**Genomic context**

Location: 7q31.1

See FOXP2 in [Genome Data Viewer](#) [Map Viewer](#)

Exon count: 24

Annotation release	Status	Assembly	Chr	Location
<a href="#">108</a>	current	GRCh38.p7 ( <a href="#">GCF_000001405.33</a> )	7	NC_000007.14 (114086310..114693772)
<a href="#">105</a>	previous assembly	GRCh37.p13 ( <a href="#">GCF_000001405.25</a> )	7	NC_000007.13 (113726365..114333827)



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# Homo sapiens chromosome 7, GRCh38.p7 Primary Assembly

NCBI Reference Sequence: NC\_000007.14

[GenBank](#) [Graphics](#)

>NC\_000007.14:114086310-114693772 Homo sapiens chromosome 7, GRCh38.p7 Primary Assembly

```

AGACAGCGCGAGCCTCCGAGAAAGCGCGAGACACGCCGGCGCGTGACGCTCCGCGGCCGCCGCTTCGCCC
TAGCTCTAGCCCCGCCACCCCGACGCCGCCCGGAACGCCGCCCGGTTATTTATGCGGCGGCCGCG
TCCGCTGGCTGCGGCTTCCTCGGCCCCCCCTCCCCGGGCGCGCCCCCACTCGCGGCAGCAGCTGCC
GGACTCGCGCGTGGGTGTGTTGTTGGGGGCTTGCCTCGCCGCGCGGGTGCCACCTCCCGGGACGCT
GCCACGGCGTCCCCGGTGGGTAAGTTTCTTGGCCCTCACTCTGGCGGCTACACCTCCGCACCCAC
CCTGTCCCAGCCACCTCCACGCTGGGCCGAGCTGCGACTTACTCTGCTCCGCGCTCCTCCCGGGTGGC
GACAAAGTTTCGCCCAAAGGCAGCGCCCTGCTTGCCGGGGCAGTGTGACATGTGTCAAATTTGGGCTC
GGCGTTGGGGTGCATTCCGGACCCAGCATCACCACTTGTGTTCTTTTCATTTGCTTTGTGATGGGG
GAAAAAGGGGTGGAGAGGGGAGATTGCTGTTGGCTCACGGATGATTTTTAGTTTGGATAATGCAAATG
TTGCTTCGTCCGGAGAGACCTCGGCTGGAGGAGAATGTGTGCGAAACACCAGATGTGTGTTGTTACT
CTCTCTTTTTAATTGTTGTTGCTTTTCCCTTCTCCCCGACCCCAACCTCGGGAGGAG
AAACAACAGTAAAAACATCTGGCGGTTAGAAGCACACACTTATTGATCCAAGTCTGACCTTTATTAC
TCAGTTGGGCAAGTGCACGCTTCTCGCGCTAAGTTGGGCACTTCAGCGTTCATCTCAGAAGTACTTCTC

```

**Change region shown**

Whole sequence

Selected region

from:  to:

**Customize view**

## Analyze this sequence

- Run BLAST
- Pick Primers
- Highlight Sequence Features
- Find in this Sequence

▶ **NCBI/Primer-BLAST: Finding primers specific to your PCR template (using Primer3 and BLAST).**


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### PCR Template

Enter accession, gi, or FASTA sequence (A refseq record is preferred)  [Clear](#)

```
TACAATTTTGATTTATTTGTATCTATATATAAGAGTGGGTATAAAATATATAAACTGTGATCGTTAGG
GTCATACAATTCATATCAGTTTTAGAGCTTTTTAACTTACAATTTGATTTATTTGTATCTATATTA
AAGAGTGGGTATAAAATCTGTACACTGTGATCATTAGGGCCAACAATTCATATCAGCTTAGAGCTT
TTTAAAGGCCCTTTCTCTGGCTGCTTAACTTCTTCTTTTATTTATTGCTTAACTAGACTCATA
TTCAGTCGTGAGTCTAGTTAAATGCTTCTTACGACACACTATTTTCATTGTTATCTAAAATTAAGTGA
```


Range


	From	To	
Forward primer	<input type="text"/>	<input type="text"/>	 <a href="#">Clear</a>
Reverse primer	<input type="text"/>	<input type="text"/>	

Or, upload FASTA file

No file chosen

### Primer Parameters

Use my own forward primer (5'→3' on plus strand)   [Clear](#)

Use my own reverse primer (5'→3' on minus strand)   [Clear](#)

	Min	Max
PCR product size	<input type="text" value="70"/>	<input type="text" value="1000"/>

# of primers to return

	Min	Opt	Max	Max T <sub>m</sub> difference
Primer melting temperatures (T <sub>m</sub> )	<input type="text" value="57.0"/>	<input type="text" value="60.0"/>	<input type="text" value="63.0"/>	<input type="text" value="3"/> 

NCBI/Primer-BLAST : results: Job id=urBlqNmz1Bvzclckz0TmFrVf9ySYTOw5mQ [more...](#)

**Input PCR template**

Range 1 - 2870

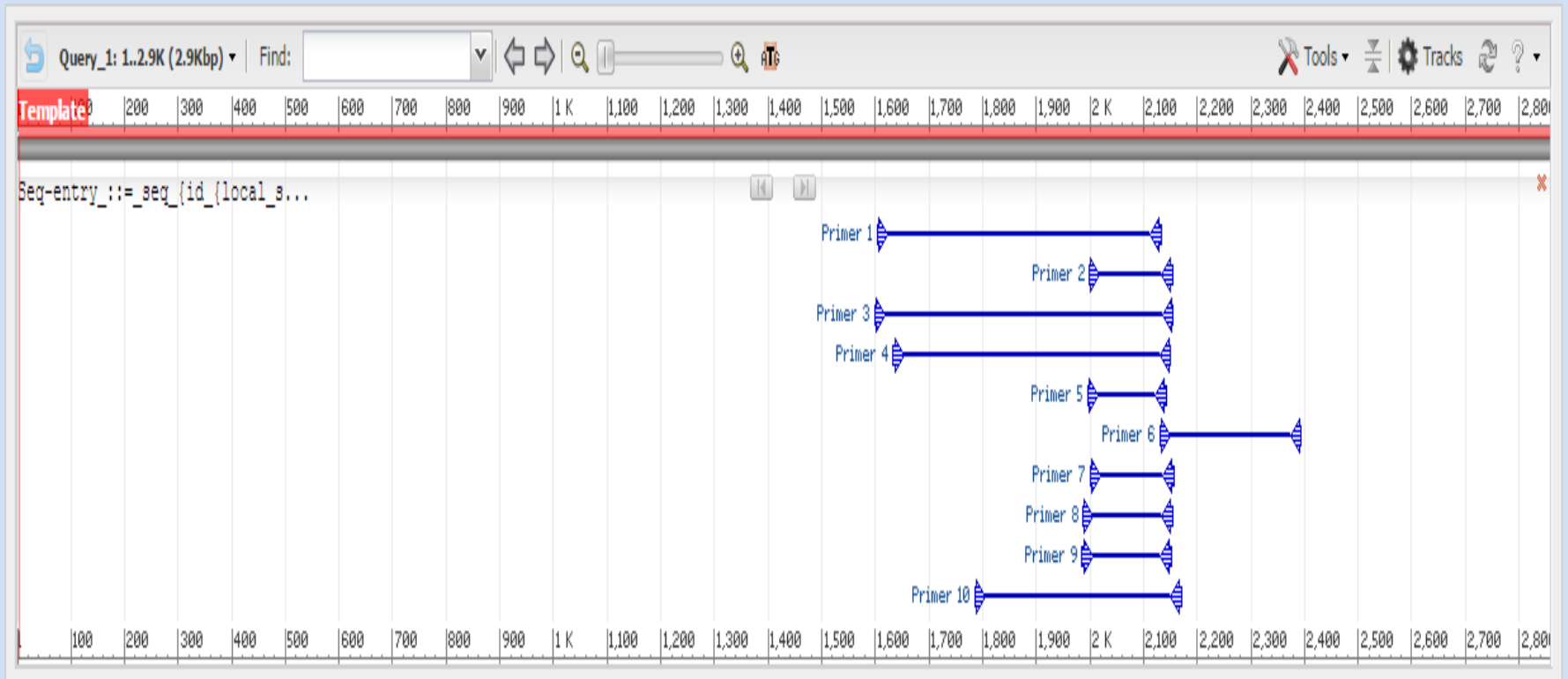
**Specificity of primers**

Primer pairs are specific to input template as no other targets were found in selected database: Refseq mRNA (Organism limited to Homo sapiens)

**Other reports**

[Search Summary](#)

## Graphical view of primer pairs



# Detailed primer reports

## Primer pair 1

	Sequence (5'→3')	Template strand	Length	Start	Stop	Tm	GC%	Self complementarity	Self 3' complementarity
Forward primer	AGTCTGGGGATTGGAAGCG	Plus	20	1604	1623	60.04	55.00	3.00	2.00
Reverse primer	CCCTTGGGCTTACCTGCTAC	Minus	20	2132	2113	60.11	60.00	4.00	1.00
Product length	529								

## Primer pair 2

	Sequence (5'→3')	Template strand	Length	Start	Stop	Tm	GC%	Self complementarity	Self 3' complementarity
Forward primer	GAAGGACCTGTGGGAGTGTG	Plus	20	1999	2018	59.96	60.00	5.00	0.00
Reverse primer	TGACAACAGGGCGTCACTTT	Minus	20	2154	2135	60.11	50.00	4.00	2.00
Product length	156								

## Primer pair 3

	Sequence (5'→3')	Template strand	Length	Start	Stop	Tm	GC%	Self complementarity	Self 3' complementarity
Forward primer	GTGCAAGTCTGGGGATTGGA	Plus	20	1599	1618	59.96	55.00	4.00	2.00
Reverse primer	TTGACAACAGGGCGTCACTT	Minus	20	2155	2136	60.11	50.00	5.00	0.00
Product length	557								

## Primer pair 4

	Sequence (5'→3')	Template strand	Length	Start	Stop	Tm	GC%	Self complementarity	Self 3' complementarity
Forward primer	GCGTTTTGCACAGGGATGAG	Plus	20	1634	1653	60.11	55.00	4.00	0.00
Reverse primer	AACAGGGCGTCACTTCTCC	Minus	20	2150	2131	60.25	55.00	3.00	1.00
Product length	517								


## Primer pair 5

	Sequence (5'→3')	Template strand	Length	Start	Stop	Tm	GC%	Self complementarity	Self 3' complementarity
Forward primer	TACGAAGGACCTGTGGGAGT	Plus	20	1996	2015	59.89	55.00	5.00	1.00
Reverse primer	CTCAGTTTCTGGGCTTCTCC	Minus	20	2149	2130	59.99	55.00	4.00	0.00

## Standard Nucleotide BLAST

[blastn](#) [blastp](#) [blastx](#) [tblastn](#) [tblastx](#)

## Enter Query Sequence

BLASTN programs search nucleotide databases using a nucleotide query. [more...](#)[Reset page](#) [Bookmark](#)Enter accession number(s), gi(s), or FASTA sequence(s) [Clear](#)Query subrange 

AGTCTGGGGATTGGAAAGCG

From



To

Or, upload file

Choose File

No file chosen 

Job Title

Enter a descriptive title for your BLAST search  Align two or more sequences 


## Choose Search Set

Database

 Human genomic + transcript  Mouse genomic + transcript  Others (nr etc.):Nucleotide collection (nr/nt) 

Organism

Optional

  Exclude Enter organism common name, binomial, or tax id. Only 20 top taxa will be shown 

Exclude

Optional

 Models (XM/XP)  Uncultured/environmental sample sequences

Limit to

 Sequences from type material

## BLAST Results

ⓘ Your search parameters were adjusted to search for a short input sequence.

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## Nucleotide Sequence (20 letters)

RID [6ZX9J7R9014](#) (Expires on 01-08 02:51 am)

Query ID [lcl|Query\\_176353](#)

Description None

Molecule type nucleic acid

Query Length 20

Database Name nr

Description Nucleotide collection (nt)

Program BLASTN 2.6.0+ [▶ Citation](#)

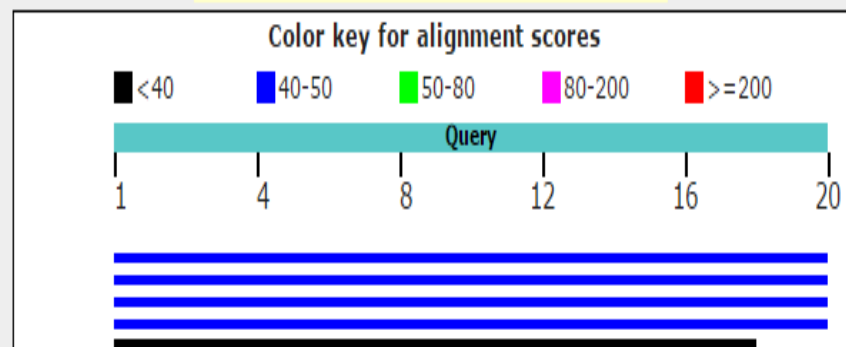
Other reports: [▶ Search Summary](#) [\[Taxonomy reports\]](#) [\[Distance tree of results\]](#)

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## Graphic Summary

## Distribution of the top 127 Blast Hits on 100 subject sequences ⓘ

Mouse over to see the title, click to show alignments



PREDICTED: Homo sapiens forkhead box P2 (FOXP2), transcript variant X1, mRNA

Sequence ID: [XM\\_017012801.1](#) Length: 7035 Number of Matches: 1

Range 1: 219 to 238 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
40.1 bits(20)	0.22	20/20(100%)	0/20(0%)	Plus/Plus

```
Query 1  AGTCTGGGGATTGGAAAGCG 20
          |||
Sbjct 219 AGTCTGGGGATTGGAAAGCG 238
```

### Related Information

Homo sapiens forkhead box P2 (FOXP2), RefSeqGene on chromosome 7

Sequence ID: [NG\\_007491.2](#) Length: 614463 Number of Matches: 1

Range 1: 6604 to 6623 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
40.1 bits(20)	0.22	20/20(100%)	0/20(0%)	Plus/Plus

```
Query 1  AGTCTGGGGATTGGAAAGCG 20
          |||
Sbjct 6604 AGTCTGGGGATTGGAAAGCG 6623
```

### Related Information

[Map Viewer](#) - aligned genomic context

Pan troglodytes BAC clone RP43-39F11 from chromosome 7, complete sequence

Sequence ID: [AC145868.2](#) Length: 209787 Number of Matches: 1

Range 1: 184048 to 184067 [GenBank](#) [Graphics](#)

▼ Next Match ▲ Previous Match

Score	Expect	Identities	Gaps	Strand
40.1 bits(20)	0.22	20/20(100%)	0/20(0%)	Plus/Minus

```
Query 1  AGTCTGGGGATTGGAAAGCG 20
          |||
Sbjct 184067 AGTCTGGGGATTGGAAAGCG 184048
```

### Related Information

[Map Viewer](#) - aligned genomic context



Select the [Task](#) for primer selection

Paste source sequence below (5'→3', string of ACGTNacgtn -- other letters treated as N -- numbers and blanks ignored). FASTA format ok. Please N-out undesirable sequence (vector, ALUs, LINEs, etc.) or use a [Mispriming Library \(repeat library\)](#)

```
TCAGTATATCCCAAGTACTGGAATTTTAAATGTAGTGTGGTTTAAATGTTGTTAAACACAATTAATT
TACAATTTGATTTATTTGTATCTATATTAAGAGTGGGTATAAAATATATAAACTGTGATCGTTAGG
GTCATACAATTCATATCAGTTTTAGAGCTTTTTAACTACAATTTTGATTTATTGTATCTATATTA
AAGAGTGGGTATAAAATCTGTCACTGTGATCATTAGGGCCAAACAATTCATATCAGCTTAGAGCTT
TTAAAGGCCCTTTCTCTTGCTGCTTAACTCTTTCCTTTATTTTATTGCCTAACTAGACTCATA
TTCAGTCGTCAGTCTAGTAAATGCTTCTTACGACACACTATTCATTGTTATCTAAAATTAAGTGA
```

Pick left primer, or use left primer below

Pick hybridization probe (internal oligo), or use oligo below

Pick right primer, or use right primer below (5' to 3' on opposite strand)




[Sequence Id](#)  A string to identify your output.

[Targets](#)  E.g. 50,2 requires primers to surround the 2 bases at positions 50 and 51. Or mark the [source sequence](#) with [ and ]: e.g. ...ATCT[CCCC]TCAT.. means that primers must flank the central CCCC.

[Overlap Junction List](#)  E.g. 27 requires one primer to overlap the junction between positions 27 and 28. Or mark the [source sequence](#) with -: e.g. ...ATCTAC-TGTCAT.. means that primers must overlap the junction between the C and T.

[Excluded Regions](#)  E.g. 401,7 68,3 forbids selection of primers in the 7 bases starting at 401 and the 3 bases at 68. Or mark the [source sequence](#) with < and >: e.g. ...ATCT<CCCC>TCAT.. forbids primers in the central CCCC.

[Pair OK Region List](#)  See manual for help.

[Included Region](#)  E.g. 20,400: only pick primers in the 400 base region starting at position 20. Or use { and } in the [source sequence](#) to mark the beginning and end of the included region: e.g. in ATC(TTC TCT)AT the included region is TTC TCT

# Primer3 Output

---

No mispriming library specified

No internal oligo mishyb library specified

Using 1-based sequence positions

OLIGO	<u>start</u>	<u>len</u>	<u>tm</u>	<u>gc%</u>	<u>any_th</u>	<u>3'_th</u>	<u>hairpin</u>	<u>seq</u>
LEFT PRIMER	956	20	59.01	55.00	5.24	0.00	0.00	AAGAAACTCCTGGGCCCTAC
RIGHT PRIMER	1143	20	59.03	55.00	0.00	0.00	0.00	GATTCACCCCAAACCCACC
INTERNAL OLIGO	988	20	60.02	50.00	19.67	7.48	39.99	ATAACCGTGACAGGGATGA

SEQUENCE SIZE: 2870

INCLUDED REGION SIZE: 2870

PRODUCT SIZE: 188, PAIR ANY\_TH COMPL: 0.00, PAIR 3'\_TH COMPL: 0.00

```
1 AGACAGCGCGAGCCTCCGAGAAAGCGCGAGACACGCCGGCGCGTGCAGCTCCGCGGCCGC
61 CGCTTCGCCCTAGCTCTAGCCCCGCGCCACCCGCAGCCCGCCCGCGAACGCCCGCCCCGG
121 TTATTTATGCGGCGGCCGCGTCCGCTGGCTGCGGCTTCTCGGCCCCCCCCCTCCCCGGG
181 CGCGCCCCCACTCGCGGCAGCAGCTGCCCGGACTCGCGCGTGGGTGTGTTGTTGGGGG
241 CTTCTGCCTCGCCGCCGCGGGTGCCACCTCCCGGGACGCTGCCACGCGTCCCCGGTCCG
301 CGGTAAGTTTCTTGGCCCTCACTCTGGCGGCTACACCTCCGCACCCACCCCTGTCCCAG
361 CCACCTCCACGCTGGGCCGAGCTGCGACTTTACTCTGCTCCGCGCTCCTCCCGCGGTGGC
```



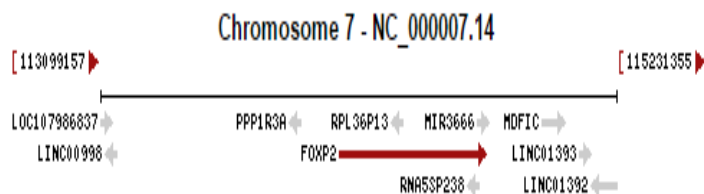
## Genomic context

Location: 7q31.1

See FOXP2 in [Genome Data Viewer](#) [Map Viewer](#)

Exon count: 24

Annotation release	Status	Assembly	Chr	Location
<a href="#">108</a>	current	GRCh38.p7 ( <a href="#">GCF_000001405.33</a> )	7	NC_000007.14 (114086310..114693772)
<a href="#">105</a>	previous assembly	GRCh37.p13 ( <a href="#">GCF_000001405.25</a> )	7	NC_000007.13 (113726365..114333827)

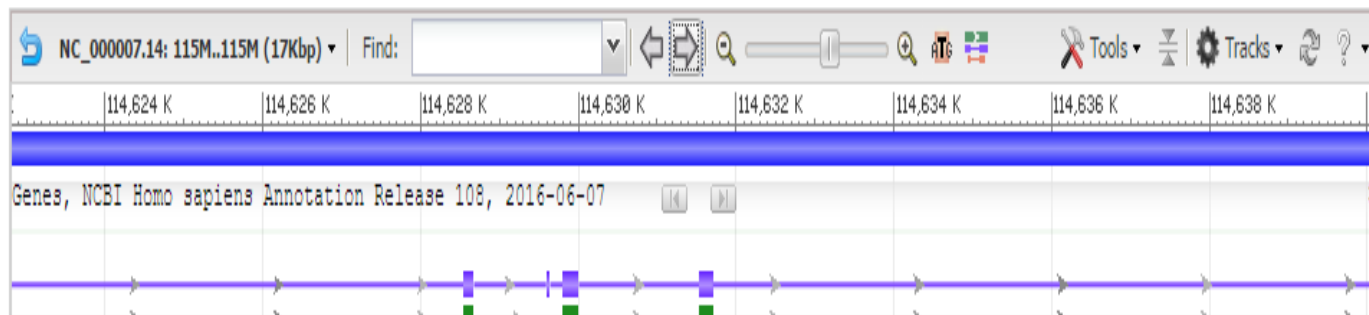


## Genomic regions, transcripts, and products

Go to [reference sequence details](#)

Genomic Sequence: [NC\\_000007.14](#) Chromosome 7 Reference GRCh38.p7 Primary Assembly

Go to nucleotide: [Graphics](#) [FASTA](#) [GenBank](#)



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[Variation Viewer \(GRCh37.p13\)](#)

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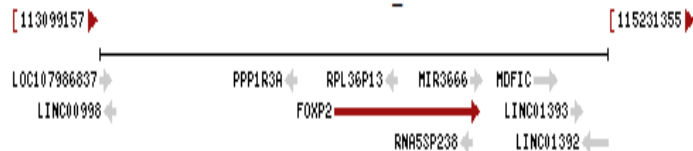
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# Chromosome 7 - NC\_000007.14



## Genomic regions, transcripts, and products

Genomic Sequence:

Go to [reference sequence details](#)

Go to nucleotide: [Graphics](#) [FASTA](#) [GenBank](#)

NC\_000007.14: 115M..115M (39bp) Find:

114,631,520 114,631,530 114,631,540 114,631,550

C T G A T A C C A G C A G C A G C A G C A G C A G C A A C A G C A A  
G A C T A T G G T C G T C G T C G T C G T C G T C G T T G T C G T

Genes, NCBI Homo sapiens Annotation Release 108, 2016-06-07

NR\_093766.1  
NR\_093767.1

Genes, Ensembl release 87

dbSNP Build 149 (Homo sapiens Annotation Release 108) all data

## Related information

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[dbVar](#)

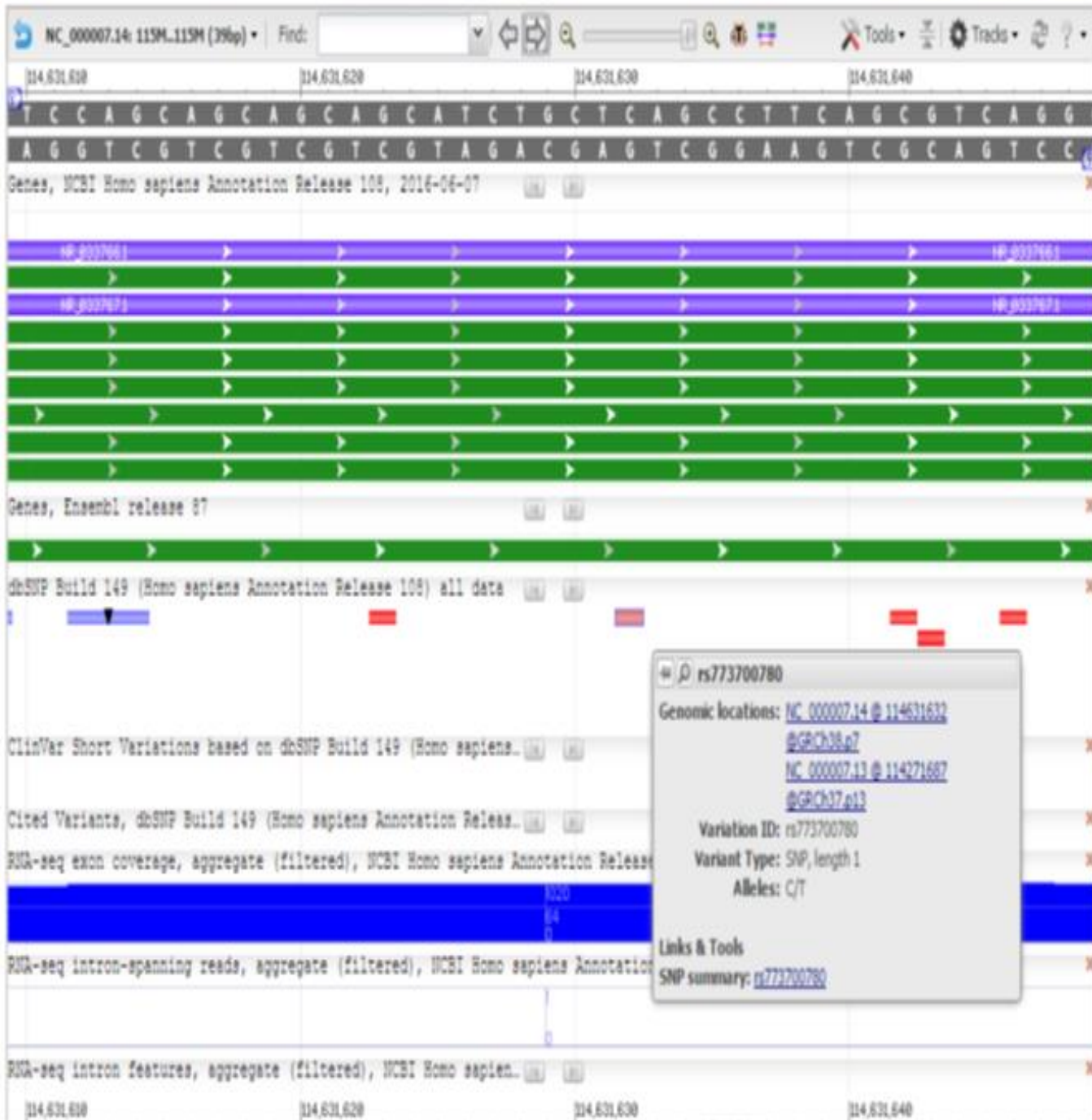
[EST](#)

[Full text in PMC](#)

[Full text in PMC\\_nucleotide](#)

[Gene neighbors](#)





**rs773700780**

Genomic locations: [NC\\_000007.14 @ 114631632 @GRCh38.p7](#)  
[NC\\_000007.13 @ 114271687 @GRCh37.p13](#)

Variation ID: rs773700780  
 Variant Type: SNP, length 1  
 Alleles: C/T

Links & Tools  
 SNP summary: [rs773700780](#)

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AACTCCAGCAGCAGCAGCATCTGCT

Y(C/T)

AGCCTTCAGC GTCAGGGACTCATCT



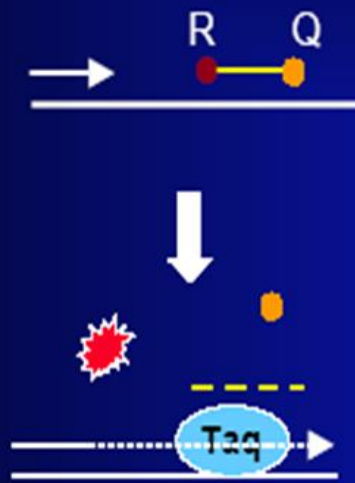
# Detection in real time PCR

## Methods of fluorescence detection

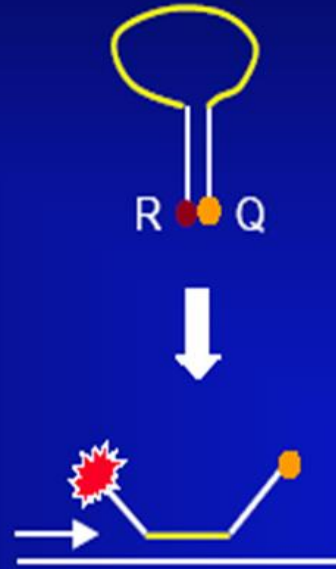
SYBR Green



Taqman



Molecular  
Beacons



Light  
Cycler



# SYBR<sup>®</sup> green

SYBR Green I fluoresces only when bound to dsDNA.

Denature



Anneal



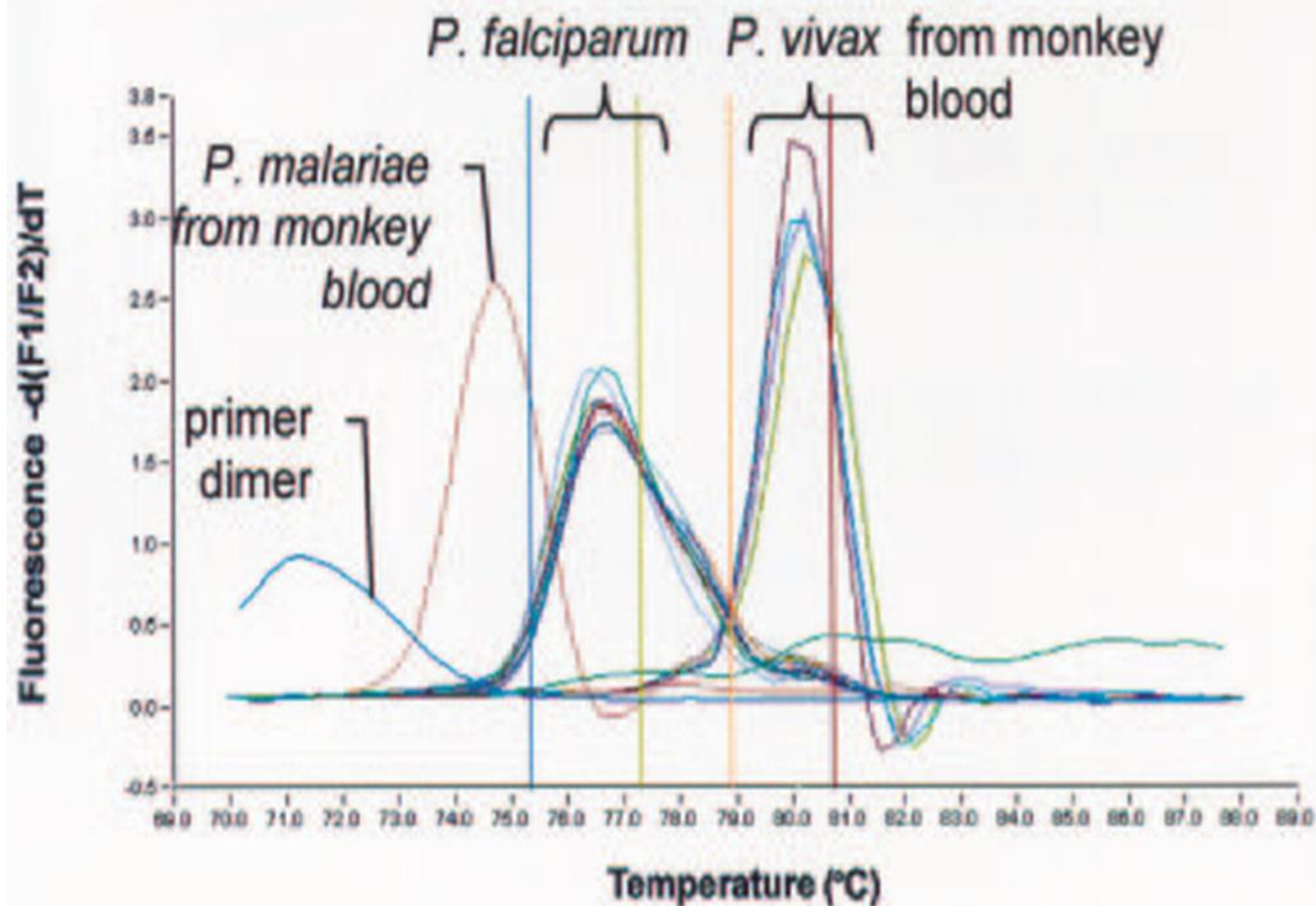
Extend



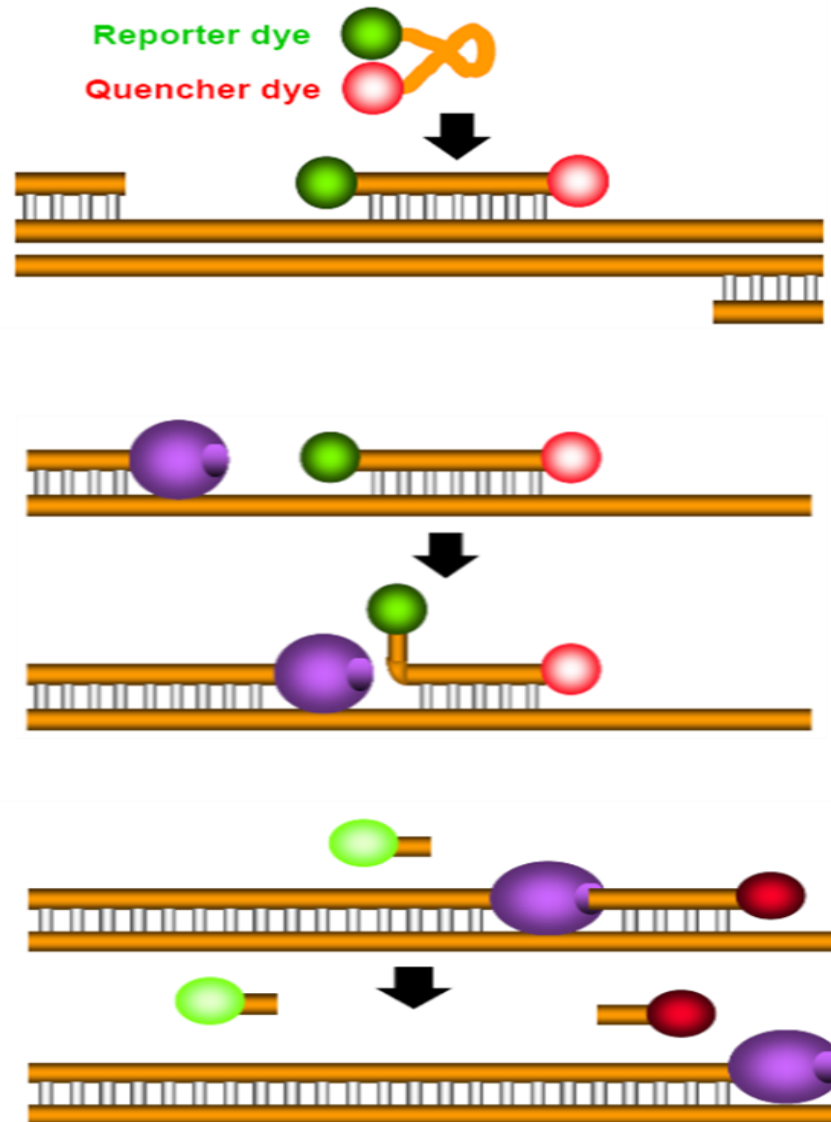
\* Pros: relatively cheap, doesn't require probe design

\* Cons: nonspecificity can lead to false positives, not attuned for complex protocols

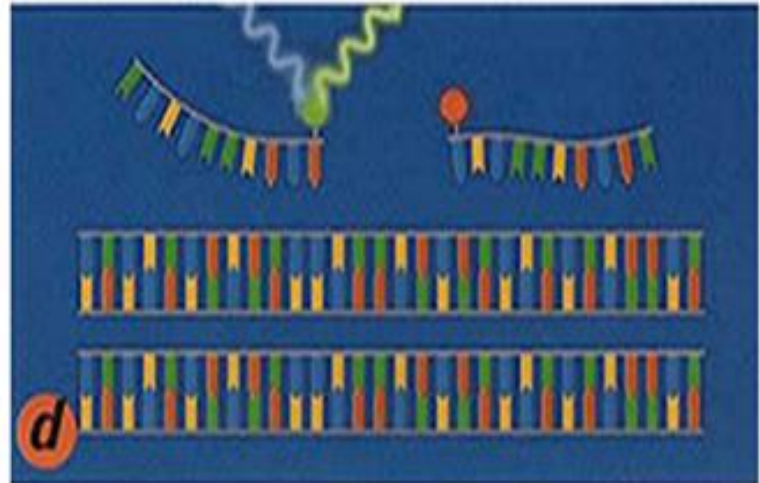
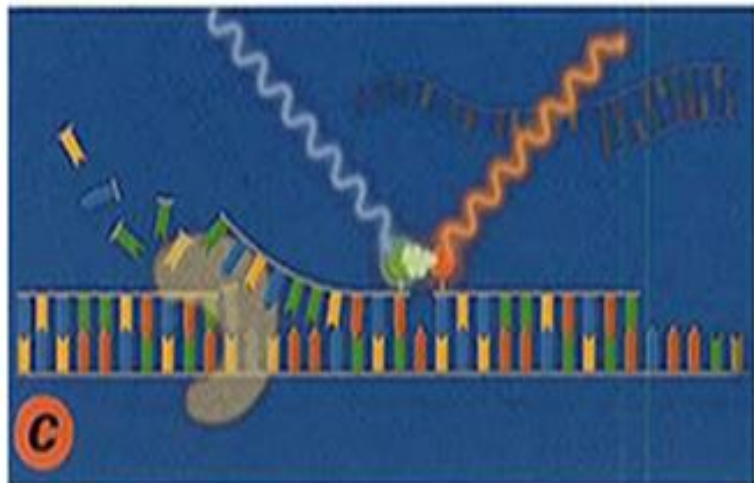
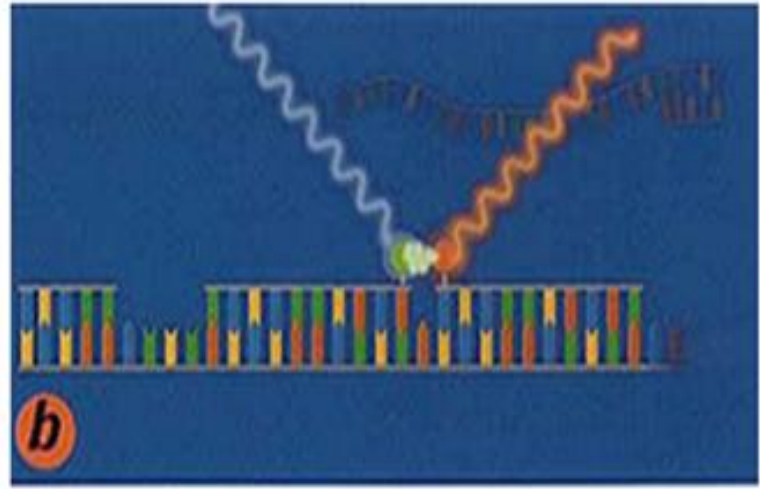
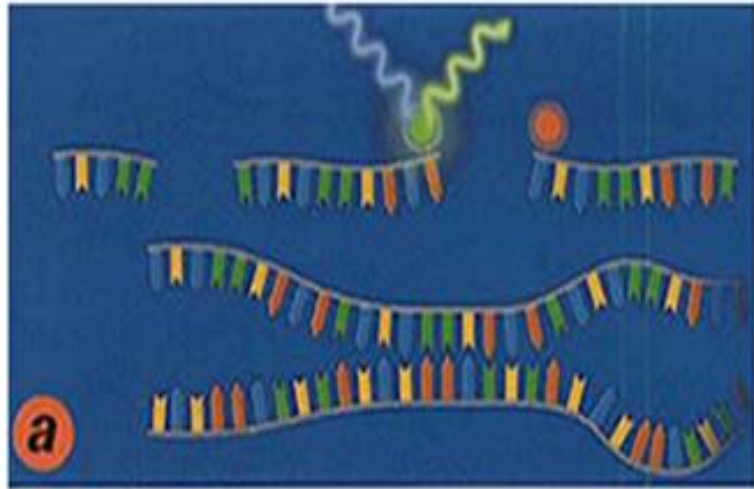




# Taqman



# Hybridization probes technique



Thank You!

